

A comparative Analysis of Antioxidant capacity of aqueous and methanolic leaf extracts of *Scoparia dulcis* and *Schleichera oleosa*

Sophy Jose*, M. P. Sinha

* Corresponding author: Sophy Jose, E-mail: josesophy35@gmail.com



*Department of Zoology, Ranchi University, Ranchi-834008, Jharkhand, India.

Abstract

In this study a comparative analysis of total anti-oxidant capacity of aqueous and methanolic leaf extracts of *Scoparia dulcis* and *Schleichera oleosa* was done by estimation of DPPH radical, hydroxyl radical, superoxide anion radical and nitric oxide radical scavenging activity. The methanolic extract (M.E) of *S.oleosa* showed a maximum concentration of total phenolics and flavanoids up to 20% and 168.89 % and aqueous extract (A.E) showed a maximum concentration of total phenolics and flavanoids up to 4.90% and 9.33% respectively. The M.E of *S.dulcis* showed a maximum concentration of total phenolics and flavanoids up to 6.21% and 9.24 % and A.E 46.22% and 14.67% respectively. Free radical scavenging activity was observed in both the M.E and A.E. For *S.dulcis* the % of free radical activity in M.E and A.E in 100µg concentration was 2.98% and 15.53% respectively. M.E and A.E of *S.oleosa* showed 54.94% and 5.93 % free radical scavenging activity correspondingly. Hydroxyl scavenging activity for 100µg of *S.dulcis* extracts were 17.92% in M.E and 11.39% in A.E where as for *S.oleosa* extracts it was 35.83% and 8.19% in M.E and A.E respectively. *S.dulcis* showed only 4.03% and 3.23% scavenging of superoxide anion radical and *S.oleosa* showed superoxide anion scavenging up to 22.58% in M.E and 3.23% in A.E. Nitric oxide scavenging activity was 5.71% in M.E of *S.dulcis* and 5.24% for A.E where as *S.oleosa* showed 33.81% nitric oxide scavenging activity in M.E and 31.43 % in A.E. The presence of phenol, flavonoid and total antioxidant in both the extract justifies the antioxidant potential of the plants which brings about their free radicals scavenging potential. Thus we conclude that the antioxidant activities may be due to the cumulative effect of the phytochemicals present in the plant which genuinely designate them as free radical scavengers.

Key words: Extracts, Total phenolic compounds, Total flavonoids, NO radical, DPPH radical, hydroxyl radical, superoxide anion radical

Introduction. Energy to fuel biological processes is produced by oxidation in many living organisms. However, there are many reactive oxygen species and free radicals that are associated or formed as a result of the oxidation process. These reactive species often cause cell death and are involved in other degenerative processes associated with ageing (Halliwell and Gutteridge 2003). Reactive oxygen species (ROS) along with free radicals are also found to play a role in functional changes associated with diseases like cancer, rheumatoid arthritis, cirrhosis etc. Exogenous chemical and endogenous metabolic processes in the human body or in the digestive system might produce highly reactive free radicals, especially oxygen derived radicals, which are capable of oxidizing biomolecules, resulting in cell death and tissue damage. (Mau *et al.*, 2001) Cells are equipped with enzymes like superoxide dismutase, catalase and also chemicals like vitamin E, vitamin C, polyphenols, carotinoids and glutathione (Niki *et al.*, 1994) to neutralize the damage caused by the radicals. These are endogenous antioxidants produced within the body to scavenge free radicals are not adequate to remove them fully and to maintain a balance (Jose, *et al.*, 2016). The use of synthetic antioxidants such as

butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), Propylgallate (PG) and butylated hydroquinone have often been implicated to achieve immediate result, recent data indicates that these synthetic antioxidants could have carcinogenic effects thus fuelling an intense search for newer and efficient antioxidants (Yevgenia *et al.*, 2013). Antioxidant containing natural foods can however be used to reduce the oxidative damage (Mau *et al.* 2001; Gülçin *et al.*, 2002).

Scoparia dulcis commonly called as sweet broom weed is a perennial herb, widely distributed in tropical and subtropical regions. Traditionally it has been used as remedies for varieties of ailments like stomach troubles, hypertension, diabetes, inflammation, bronchitis, hemorrhoids, hepatitis, analgesic, diuretic, antipyretic and cytotoxicity (Freire *et al.*, 1993; Hayashi *et al.*, 1993; Ahsan *et al.*, 2003).

Schleichera is a genus of plants in the soapberry family, *Sapindaceae* and is generally found in Indian subcontinent in mixed deciduous forests and in Southeast Asia. It is used in traditional medicine systems and its leaf, seed, oil and bark are used for curing itch, burns, acne pain in the back and

loins etc. and it promotes hair growth, treats rheumatism, head ached, skin diseases, malarial fever and is prophylactic against cholera [Palanuvej *et al.*, 2008].

Scoparia dulcis and *Schleichera oleosa* have high medicinal value and are easily available common plants which can be used in anti-oxidant therapy as well as other curative practice (Jose and Sinha. 2017). Thus, showing the potential medicinal efficacy of each extracted compounds present in the plant, permits not only the demonstration of their physiological activity but also facilitates pharmacological studies leading to synthesis of more potent drugs with reduced toxicity (Manna and Abalaka, 2000). Hence, it is important to compare the antioxidant efficacy of *S. dulcis* and *S. oleosa*. Therefore present study was undertaken to compare the antioxidant capacity of aqueous and methanolic leaf extracts of *Scoparia dulcis* and *Schleichera oleosa*.

MATERIALS AND METHODS

Collection of plant materials

The fresh mature leaves were collected, dried in shade under room temperature for six to seven days and then crushed into coarse powder using electric grinder. The powder was sieved to get fine powder using fine plastic sieve which was stored in air tight bottle in the laboratory until required.

Extract preparation

50g of the powder was subjected to extraction by soxhlet using methanol and distilled water separately. The extracts obtained were filtered, concentrated after dryness in rotary flash evaporator maintained at 45°C., percentage yield of each extract was calculated and the dried extracts were stored in air tight containers at room temperature for further studies.

Phytochemical analyses

Freshly prepared extracts of the powdered leaves were subjected to phytochemical analyses to find the presence of the following phyto constituents such as flavanoids, alkaloids, carbohydrates, glycosides, polysaccharides, tannins, saponins, steroids, proteins, lipids, oils by standard methods.[Trease and Evans, 2002; Sofowara, 2008].

Estimation of total phenolics

Known amount of sample were pipetted out in series of test tubes and volume was made up to 3 ml with distilled water. Folin-Ciocalteu reagent (0.5ml) was added to each tube and incubated for 3 min. at room temperature Sodium carbonate (20%; 2ml) solution was added, mixed thoroughly and the tubes were incubated for 1 min. in boiling water bath. Absorbance was measured at 650nm against a reagent blank. Standard curve using different concentrations of standard phenolic -catechol was prepared. From the standard curve, concentration of phenols in the test samples was determined and expressed as mg of catechol equivalent. (Malick, *et al.*, 1980) The values are furnished in Fig.1.

Estimation of flavonoids

Known volume of samples was pipetted out in series of test tubes and volume was made up to 0.5 ml with distilled water. Sodium nitrite (5%; 0.03ml) was added to each tube and incubated for 5 min. at room temperature. Aluminium chloride solution (10%; 0.06ml) solution was added and incubated for 5 min. at room temperature. Sodium Hydroxide solution (1 M; 0.2ml) was added and total volume was made up to 1 ml with distilled water. Absorbance was measured at 510 nm against a reagent blank. Standard curve using different concentrations of rutin was prepared. From the standard curve, concentration of flavonoids in the test samples was determined and expressed as mg of rutin equivalent. [Helmja, *et al.*, 2007].The values are given in Fig.2.

Estimation of total antioxidant activity

The antioxidant properties of plant samples were determined by Spectrophotometric quantitation method (Prieto *et al.*,1999). Various concentrations of samples (5 µg, 50 µg, 100µg) were taken in a series of test tubes. The 1.9mL of reagent solution (0.6m Sulphuric acid, 28mm Sodium phosphate and 4mm Ammonium molybdate) was added to the test tubes. The tubes were incubated at 95°C for 90 min and allowed to cool down. The absorbance of aqueous solution of each was measured at 695nm against blank. Antioxidant capacities were expressed as equivalents of ascorbic acid. Butylated hydroxyanisole (BHA) was used as reference standard.

Estimation of free radical scavenging activity

Different concentrations (10µg, 50µg and 100µg) of samples in Dimethyl sulfoxide (DMSO), were taken in a series of test tubes. The volume was adjusted to 500µl by adding Methanol. Five millilitres of a 0.1 mm methanolic solution of 1,1-diphenyl-2-picrylhydrazyl (DPPH; from Sigma –Aldrich, Bangalore) was added to these tubes and shaken vigorously. A control without the test compound, but with an equivalent amount of methanol was maintained. The tubes were allowed to stand at RT for 20 min. The absorbance of the samples was measured at 517 nm. Butylated Hydroxy Anisole (BHA) was used as reference standard. [Kumar, *et al.*, 2008]: Free Radical scavenging activity was calculated using the following formula:

$$\% \text{ radical scavenging activity} = \frac{(\text{control OD} - \text{sample OD})}{\text{Control OD}} \times 100.$$

Estimation of hydroxyl Radical Scavenging Activity

Various concentrations (10µg, 50µg, and 100µg) of samples in DMSO were taken in different test tubes and made up to 250µl with 0.1M phosphate buffer. One milliliter of iron-EDTA solution (0.13% ferrous ammonium sulfate and 0.26% EDTA), 0.5 ml of EDTA (0.018%), and 1 ml of Dimethyl sulphoxide (0.85% v/v in 0.1 M phosphate buffer, pH 7.4) were added to these tubes, and the reaction was initiated by adding 0.5 ml of 0.22% ascorbic acid. These reaction mixtures were incubated at room temperature for 15 min. The reaction was terminated by the addition of 1 ml of ice-cold TCA (17.5% w/v). Three milliliters of Nash reagent (150 g of ammonium acetate, 3 ml of glacial acetic acid, and 2 ml of acetyl acetone were mixed and raised to 1 L with distilled water) was added to all of the tubes and left at room temperature for 15 min for color development. The intensity of the yellow color formed was measured spectrophotometrically at 412 nm against reagent blank. Ascorbic acid (AA) was used as reference standard. (Klein, *et al.*, 1991).

The percentage hydroxyl radical scavenging activity was calculated by the following formula:

% hydroxyl radical scavenging activity

$$= 1 - \left(\frac{\text{difference in absorbance of sample} \times 100}{\text{difference in absorbance of blank}} \right)$$

Estimation of super oxide anion Radical Scavenging Activity

Nitroblue tetrazolium (NBT; 150 µM in Tris-HCl buffer 16mM pH 8.0; 1ml) and Nicotinamide adenine dinucleotide (NADH; 234 µM in Tris-HCl buffer 16mM pH 8.0; 1ml) were mixed in a series of test tube. Various concentrations (10µg, 50µg and 1000µg) of samples and Ascorbic acid (AA) were added to these test tubes and made up to 3ml with Tris-HCl buffer (16mM; pH 8.0). Ascorbic Acid (AA) was used as reference standard for comparison. Phenazine methosulphate solution was added (40 µM; 1ml) to each test tube. The reaction mixture was incubated for 5 min at RT. A control without the test compound was maintained. The absorbance of the samples was measured at 560 nm. [Gulcin, *et al.*, 2005]. Super oxide radical scavenging activity was calculated using the following formula:

$$\% \text{ SO radical scavenging activity} = \frac{(\text{control OD} - \text{sample OD})}{\text{control OD}} \times 100.$$

Estimation of nitric oxide radical Scavenging Activity

Various concentrations (10µg, 50µg and 100µg) of samples and Butylated hydroxy anisole (BHA) were taken in different test tubes and made up to 3ml with 0.1M phosphate buffer (pH 7.2). Sodium Nitroprusside (5mM) prepared in buffered saline (pH 7.2) was added (1 ml) to each tube. The reaction mixture was incubated for 30 min at RT. A control without the test compound, but with an equivalent amount of methanol was maintained. After 30 min, 1.5 ml of above solution was mixed with 1.5 ml of Griess reagent (1% Sulphanilamide, 2% phosphoric acid and 0.1% N-1- Naphthylethylenediamine dihydrochloride). The absorbance of the samples was measured at 546 nm. [Kumar, *et al.*, 2008]. Nitric oxide radical scavenging activity was calculated using the following formula:

$$\% \text{ NO radical scavenging activity} = \frac{(\text{control OD} - \text{sample OD})}{\text{control OD}} \times 100.$$

RESULTS AND DISCUSSIONS

Phytochemical analyses

The results of the evaluation of phytochemical screening of methanolic extracts of *Schleichera oleosa* and *Scoparia dulcis* revealed the presence of carbohydrates, glycosides, polysaccharides, proteins, steroids, alkaloids, triterpenoids, tannins, lipids, oils, and flavanoids. Saponins were found to be present in the methanolic extract of *Schleichera oleosa* but were absent in both the extracts of *Scoparia dulcis*. These constituents are responsible for the curative nature of *Schleichera oleosa* against itching, head ache, malaria, skin diseases etc. which could make the plant useful for treating different ailments and having a potential of providing useful and safe drugs for human use.

Table 1. Proximate Phytochemical composition of Methanolic and aqueous extracts of *S.Oleosa* and *S.dulcis*

Phytochemicals	<i>S. oleosa</i>		<i>S.dulcis</i>	
	Methanolic	Aqueous	Methanolic	Aqueous
Carbohydrates	+	+	+	+
Glycosides	+	+	+	+
Polysaccharides	+	-	-	-
Proteins	+	+	+	+
Alkaloids	+	+	+	+
Steroids	+	+	+	+
Triterpenes	+	-	+	-
Flavanoids	+	-	+	+
Tannins	+	-	+	-
Lipid	-	+	+	+
Oils	+	+	+	-
Saponins	-	+	-	-

Total phenolic, flavonoid contents and total antioxidant activity

As reported by Mahakunakorn *et al.*, (2004) these phenolics and flavonoids compounds present in extracts are believed to intercept the free-radical chain of oxidation and donate hydrogen from the phenolic hydroxyl groups, thereby forming stable free radicals, which do not initiate or propagate further oxidation.

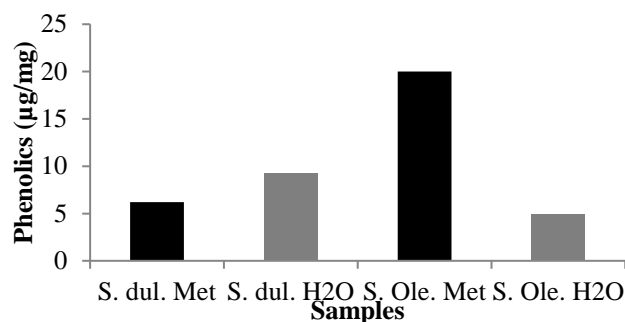


Fig.1. Total phenolics contents of *S.oleosa* and *S.dulcis*

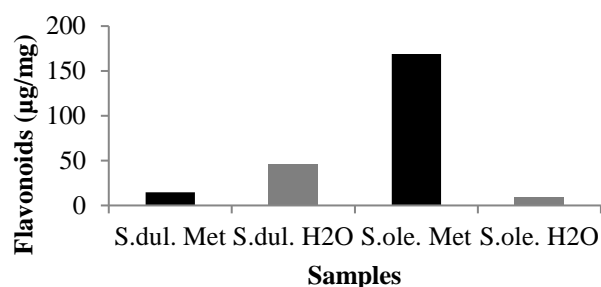


Fig. 2. Flavonoid content of leaf extracts of *S.dulcis* and *S.oleosa*

Total antioxidant capacity

DPPH, hydroxyl, superoxide and nitric oxide radical scavenging activity are the most widely used methods for screening the antioxidant activity of plant extract. The extracts displayed a creditable DPPH scavenging (Fig-4) hydroxyl radical scavenging (Fig-5) superoxide ion scavenging (Fig-6) and nitric oxide scavenging activity (Fig-7) for their respective extract. However, in the entire four tests methanol extract displayed better scavenging activity than aqueous extract for this study. These findings suggest that the plant extracts could have contained phytochemicals that is capable of donating hydrogen to a free radical in order to remove the odd electron which is responsible for the radical's reactivity.

Table.2.Total anti-oxidant property analysis of leaf extracts of *S.dulcis* and *S.oleosa*.

Concentration (µg)	<i>S.dul.</i> Met	<i>S.dul.</i> H2O	<i>S.ole.</i> Met	<i>S.ole.</i> H2O	BH A
10	2.9	6.05	7.8	1.85	5
50	7.4	13.1	19.35	5.15	14.5
100	12.8	19.9	25.5	7.7	28

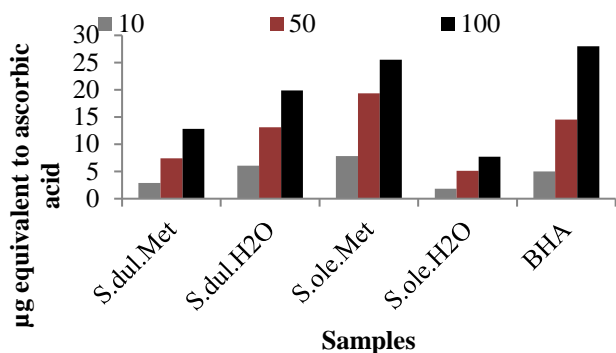


Fig.3. Showing total anti-oxidant property of extracts of *S.dulcis* and *S.oleosa*

Free radical scavenging assay

Interception of free radicals is mainly by radical scavenging, The scavengers include various antioxidants like carotenoids, flavonoids, etc., (Sies, 1996). In the present study, aqueous and methanolic extracts were found to be effective scavengers against DPPH radical. Their activities increased in a concentration dependent manner (Fig. 4). The methanolic extracts showed higher DPPH radical scavenging activity, than aqueous extracts. The DPPH radical scavenging activity of methanolic extracts of *S.oleosa* was shown to be 54.94% at 100µg/ml and of aqueous extract was 5.92 % at 100 µg/mg . For *S.dulcis* the % of free radical activity in methanolic extract and aqueous extract in 100µg concentration was 2.98% and 15.53% respectively.

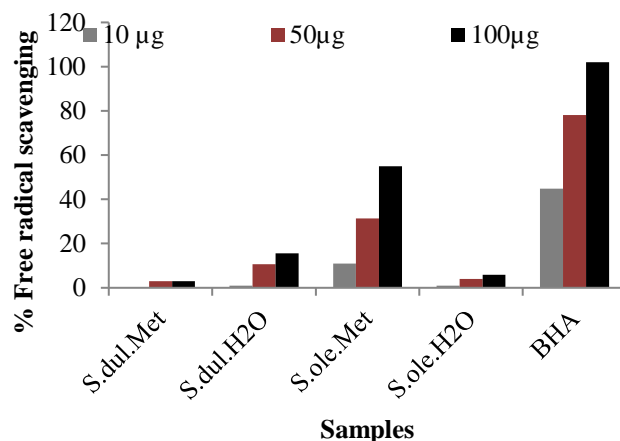


Fig4: Percentage Free radical scavenging activity of *S.dulcis* and *S.oleosa* in comparison with BHA.

Hydroxyl radical scavenging activity

Hydroxyl radicals are the major active oxygen species that cause lipid oxidation and enormous biological damage (Aurand, *et al.*, 1977, Sunil, *et al.*, 2008).The hydroxyl radical scavenging activity of the methanolic and aqueous extracts was studied (Fig. 5). Both extracts exhibited a concentration-dependent scavenging abilities for the hydroxyl radical. Methanolic extract was found to be the more powerful scavenger of the hydroxyl radical, with an inhibition of up to 35.83 % and 17.92% for *S.oleosa* and *S.dulcis* respectively at a concentration of 100µg/ml. It is worth mentioning that aqueous extract showed an inhibition of 8.19 % and 11.39% at a concentration 100µg/ml for *S.oleosa* and *S.dulcis* respectively.

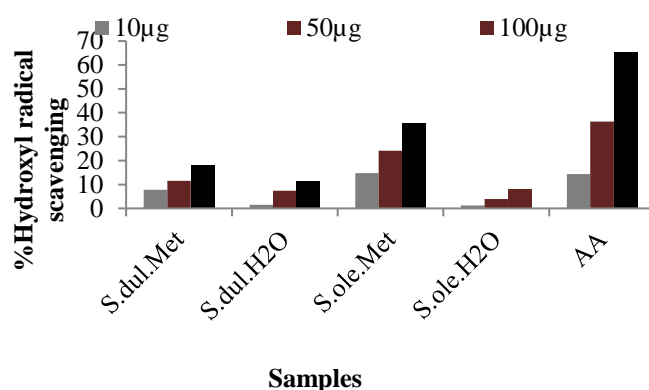


Fig.5: Percentage hydroxyl radical scavenging activity of *S.dulcis* and *S.oleosa* in comparison with Ascorbic acid

Superoxide anion scavenging activity

Earlier work on the percentage inhibition of superoxide generation by *Citrullus colocynthis* at 2500 mg mL⁻¹ concentration was found to be 71.3 ± 3.2% (p < 0.005), (Sunil *et al.*,2008).The Superoxide anion scavenging activity of the methanolic and aqueous extracts was explored (Fig. 6). Both extracts exhibited concentration-dependent scavenging abilities for the superoxide anion radical. Methanolic extract was found to be the more powerful scavenger of the superoxide anion radical, with an inhibition of up to 22.58 % at a concentration of 100µg/ ml and aqueous extract showed an inhibition of 3.23 at a concentration 100µg/ml for *S.oleosa* and *S.dulcis* showed only 4.03% and 3.23% scavenging of superoxide anion radical at a concentration of 100µg/ml for methanolic and aqueous extracts respectively

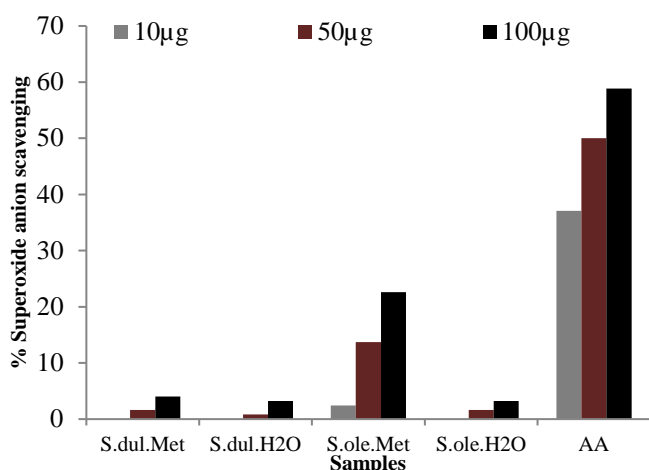


Figure.6. super-oxide anion radical scavenging activity of *S.dulcis* and *S.oleosa*

The nitric oxide scavenging activity

Earlier works have shown nitric oxide scavenging capacity of different plant parts (Wankupar, *et al.*, 2015). Methanolic and aqueous leaf extracts of *Schleichera oleosa* and *Scoparia dulcis* exhibited concentration-dependent scavenging abilities for the nitric oxide radical. M.E was found to be the more powerful scavenger of the nitric oxide radical, with an inhibition of up to 33.81 % at a concentration of 100µg/ ml and A.E showed an inhibition of 31.43% at a concentration 100µg/ml in case of *S.oleosa*. Nitric oxide scavenging activity was 5.71% in M.E of *S.dulcis* and 5.24% for A.E.

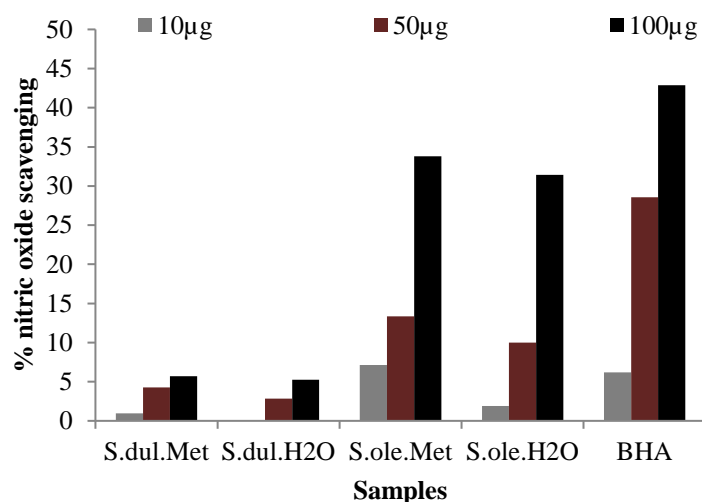


Fig.7.Nitric oxide scavenging assay of leaf extracts of *S.dulcis* and *S.oleosa*

Conclusion

Thus summarizing these results, it is evident that *S.oleosa* has better anti-oxidising capacity than *S.dulcis*. It is also evident that methanol extracts of *S. dulcis* and *S. oleosa* proved to have superior antioxidant capacity when compared to aqueous extract in this particular study and this may have resulted due to the greater extraction capacity of methanol when used as solvent. Hence, the possibility of using crude extracts of *S.oleosa* and *S.dulcis* as antioxidants would greatly reduce the need to obtain pure compounds via expensive industrial purification techniques. Further in depth study of toxicity and dosage may reveal the efficacy of the plants as alternatives to anti-oxidant therapy. On the basis of above studies, it can be concluded leaf of *Schleichera oleosa* and *Scoparia dulcis* contains several beneficial compounds such as flavonoids, phenols and tannins. Our studies presented a positive relationship between the free radical scavenging activity, the effective antioxidant capacity displayed by total phenol, flavonoids and antioxidant content in both methanol and aqueous extract which clearly justifies the therapeutic efficacy of both plant leaf extracts as an alternative to synthetic antioxidant. In conclusion, *S.oleosa* and *S.dulcis* might be agents with anti-cancer, liver protective and anti-inflammatory potentials as they show moderate antioxidant activity and should be further investigated in the future.

ACKNOWLEDGEMENT

The authors acknowledge the facilities provided by Department of Zoology, Ranchi University, Ranchi, India for carrying out the work.

Bibliography

1. **Halliwell B, Gutteridge JMC. 2003.** Free radicals in biology and medicine. Oxford University Press.
2. **Mau JL, Chao GR, Wu KT. 2001.** Antioxidant properties of methanolic extracts from several mushrooms. *J Agric Food Chem.* 49: 5461–5467.
3. **Niki E, Shimaski H, Mino M. 1994.** Antioxidantism—free radical and biological defense Tokyo. Japan: Gakkai Syuppan Center.
4. **Sophy Jose and M.P. Sinha. 2016.** Phytochemistry and antibacterial efficacy of *Schleichera oleosa* on some human pathogenic bacteria. *J. The Eco Scan Special issue, Vol. IX:* 00-00.
5. **Yevgenia Shebis, David Iluz, Yael Kinel-Tahan, Zvy Dubinsky and Yaron Yehoshua. 2013.** Natural antioxidants: function and sources. *Food Nutr Sci,* 4: 643-649.
6. **Gülçin I, Oktay M, Küfrevioğlu ÖI, Aslan A. 2002.** Determination of antioxidant activity of lichen *Cetraria islandica* (L.) Ach. *J Ethnopharmacol.* 79: 325–329
7. **Freire SMF, Emim AJS, LAPA AJ, Souccar C, Torres LMB. 1993.** Analgesic and anti-inflammatory properties of *Scoparia dulcis* L. extract and glutinol in rodents. *Phytother Res,* 7:408–414.
8. **Hayashi T, Kawasaki M, Miwa Y, Taga T, Morita N.** Antiviral agents of plant origin. III. Scopadulin, a novel tetracyclic diterpene from *Scoparia dulcis* L. *Chem Pharm Bull,* 1990; 3: 945–947.
9. **Ahsan M, Islam SKN, Gray AI, Stimson WHC.** Cytotoxic diterpenes from *Scoparia dulcis*. *J Nat Prod,* 2003; 66(7): 958-61.
10. **Palanuvej C, Vipunngeun N.** Fatty acid constituents of *Schleichera oleosa* (Lour) Oken seed oil. *J Health Res.* 2008;22:203–2.
11. **Sophy Jose and M.P. Sinha. 2017.** Study of Antibacterial Efficacy of Methanolic and Aqueous Leaf Extracts of *Scoparia dulcis* on Some Human Pathogenic Bacteria. *Int.J.Curr.Microbiol.App.Sci.* 6(1): 423-432.
12. **Manna A, Abalaka ME.** Preliminary screening of the various extracts of *Physalis angulata* (L.) for antimicrobial activities. *Spectrum J,* 2000 7(2):119-125.
13. **Trease, G. E. and Evans, W. C. 2002.** Pharmacognosy. 15th Ed. Saunders Publishers, London. p. 42.
14. **Sofowora, A. 2008.** Screening Plants for Bioactive Agents. In: Medicinal Plants and Traditional Medicinal in Africa. 3rd Ed. Spectrum Books Ltd, Sunshine House, Ibadan, Nigeria. p. 134.
15. **Mallick, C.P. and Singh, M.B. (1980).** In: *Plant Enzymology and Histoenzymology.* Kalyani Publishers. New Delhi. P 286)
16. **Helmja K., Vaher M., Gorbatošova J. and Kaljur and M. (2007).** Characterization of bioactive compounds contained in vegetables of the Solanaceae family by capillary electrophoresis. *Proc. Estonian Acad. Sci. Chem.,* 56; (4) 172–186.
17. **Prieto P., Pineda M. and Aguilar M. (1999).** Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application to the determination of vitamin E1. *Analytical Biochemistry.* 269: 337-341
18. **Kumar S., Kumar D., Manjusha, Saroha K., Singh N. and Vashishta B.(2008):** Antioxidant and free radical scavenging potential of *Citrullus colocynthis* (L.) Schrad. methanolic fruit extract, *Acta. Pharm.* 58; 215–220.
19. **Klein, S. M.; Cohen, G.; Cederbaum, A. I. (1991).** Production of formaldehyde during metabolism of dimethyl sulphoxide by hydroxyl radical generating system. *Biochemistry,* 20, 6006-6012.
20. **Gulcin I., Alici H.A. and Cesur M. (2005).** Determination of *in vitro* antioxidant and radical scavenging activities of Propofol. *Chem. Pharm. Bull.* 53(3); 281-285.
21. **P. Mahakunakorn, M. Tohda, Y. Murakami, K. Matsumoto, H. Watanabe, 2004,** Antioxidant and free radical scavenging activity of Choto-san and its related constituents, *Biol. Pharm. Bull.,* 27, pp. 38-46.
22. **Sies H. (Ed.)** Antioxidants in Disease, Mechanisms and Therapy, Academic Press, New York, 1996.
23. **L. W. Aurand, N. H. Boonme and G. G. Gidding.** Superoxide and singlet oxygen in milk lipid peroxidation, *J. Dairy Sci.* 60 (1977) 363–369.
24. **Sunil Kumar, Dinesh Kumar, Manjusha, Kamal Saroha, Nidhan Singh, Bhoodev Vashishta.** Antioxidant and free radical scavenging potential of *Citrullus colocynthis* (L.) Schrad. methanolic fruit extract. *J. Acta Pharm.* 58 (2008) 215–220
25. **Wankupar Wankhar, Sakthivel Srinivasan, Ravindran Rajan, Sheeladevi Rathinasamy.** Evaluating *in vitro* antioxidant activity and GC-MS analysis of *Scoparia dulcis* Linn (Scrophulariaceae). *J. Applied Pharmaceutical Science* Vol. 5 (07), pp. 029-034, July, 2015